



ORIGINAL ARTICLE

Susceptibility to rifaximin and other antimicrobial agents of bacteria isolated from acute gastrointestinal infections in Mexico^{☆,☆☆}



O. Novoa-Farías^a, A.C. Frati-Munari^{b,*}, M.A. Peredo^c, S. Flores-Juárez^a,
O. Novoa-García^a, J. Galicia-Tapia^a, C.E. Romero-Carpio^a

^a División de Microbiología Clínica, Unidad de Diagnóstico Microbiológico UDMSC, Mexico City, Mexico

^b Departamento de Medicina Interna, Hospital Médica Sur, Mexico City, Mexico

^c Alliance for the prudent use of antibiotics (APUA), Chapter Mexico, Mexico City, Mexico

Received 23 April 2015; accepted 14 July 2015

Available online 1 February 2016

KEYWORDS

Rifaximin;
Bacterial resistance;
Bacterial
susceptibility;
Antibiotics;
Gastroenteritis;
Small intestinal
bacterial overgrowth

Abstract

Background: Bacterial resistance may hamper the antimicrobial management of acute gastroenteritis. Bacterial susceptibility to rifaximin, an antibiotic that achieves high fecal concentrations (up to 8,000 µg/g), has not been evaluated in Mexico.

Objective: To determine the susceptibility to rifaximin and other antimicrobial agents of enteropathogenic bacteria isolated from patients with acute gastroenteritis in Mexico.

Material and methods: Bacterial strains were analyzed in stool samples from 1,000 patients with diagnosis of acute gastroenteritis. The susceptibility to rifaximin (RIF) was tested by microdilution (< 100, < 200, < 400 and < 800 µg/ml) and susceptibility to chloramphenicol (CHL), trimethoprim-sulfamethoxazole (T-S), neomycin (NEO), furazolidone (FUR), fosfomicin (FOS), ampicillin (AMP) and ciprofloxacin (CIP) was tested by agar diffusion at the concentrations recommended by the Clinical & Laboratory Standards Institute and the American Society for Microbiology.

Results: Isolated bacteria were: enteropathogenic *Escherichia coli* (*E. coli*) (EPEC) 531, *Shigella* 120, non-Typhi *Salmonella* 117, *Aeromonas spp.* 80, enterotoxigenic *E. coli* (ETEC) 54, *Yersinia enterocolitica* 20, *Campylobacter jejuni* 20, *Vibrio spp.* 20, *Plesiomonas shigelloides* 20, and enterohemorrhagic *E. coli* (EHEC O:157) 18. The overall cumulative susceptibility to RIF at <100, <200, <400, and <800 µg/ml was 70.6, 90.8, 99.3, and 100%, respectively. The overall susceptibility to each antibiotic was: AMP 32.2%, T-S 53.6%, NEO 54.1%, FUR 64.7%, CIP 67.3%,

[☆] Please cite this article as: Novoa-Farías O, Frati-Munari AC, Peredo MA, Flores-Juárez S, Novoa-García O, Galicia-Tapia J, et al. Susceptibilidad de las bacterias aisladas de infecciones gastrointestinales agudas a la rifaximina y otros agentes antimicrobianos en México. Revista de Gastroenterología de México. 2016;81:3-10.

^{☆☆} See related content at doi: <http://dx.doi.org/10.1016/j.rgmx.2016.01.001>, Remes Troche JM. Reflexiones sobre la resistencia a antibióticos y qué hacer al respecto. Rev Gastroenterol Méx. 2016;81(1):1-2.

* Corresponding author. Alfa Wassermann S.A. de C.V., Av. Insurgentes Sur 2453-501, Col. Tizapán San Ángel. Del Álvaro Obregón, México D.F., CP 01090. México. Tel.: +5481 4707.

E-mail addresses: afrazi@alfawassermann.com, afratim@hotmail.com (A.C. Frati-Munari).

PALABRAS CLAVE

Rifaximina;
Resistencia
bacteriana;
Susceptibilidad
bacteriana;
Antibióticos;
Gastroenteritis;
Sobrepoblación
bacteriana intestinal

CLO 73%, and FOS 81.3%. The susceptibility to RIF <400 and RIF <800 µg/ml was significantly greater than with the other antibiotics ($p < 0.001$).

Conclusions: Resistance of enteropathogenic bacteria to various antibiotics used in gastrointestinal infections is high. Rifaximin was active against 99-100% of these enteropathogens at reachable concentrations in the intestine with the recommended dose.

© 2015 Asociación Mexicana de Gastroenterología. Published by Masson Doyma México S.A. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Susceptibilidad de las bacterias aisladas de infecciones gastrointestinales agudas a la rifaximina y otros agentes antimicrobianos en México

Resumen

Antecedentes: La resistencia bacteriana puede dificultar el tratamiento antimicrobiano de las gastroenteritis agudas. La susceptibilidad bacteriana de los enteropatógenos a la rifaximina, un antibiótico que alcanza altas concentraciones fecales (hasta 8,000 µg/g) no se ha evaluado en México.

Objetivos: Determinar la susceptibilidad a rifaximina y a otros antimicrobianos de bacterias enteropatógenas aisladas de pacientes con gastroenteritis aguda en México.

Material y métodos: Se analizaron las cepas bacterianas en las heces de 1,000 pacientes con diagnóstico de gastroenteritis aguda. Se probó la susceptibilidad a la rifaximina (RIF) con microdilución (< 100, < 200, < 400 y < 800 µg/ml), la susceptibilidad a cloranfenicol (CLO), trimetoprim-sulfametoxazol (T-S), neomicina (NEO), furazolidona (FUR), fosfomicina (FOS), ampicilina (AMP) y ciprofloxacino (CIP) se probó por difusión-agar a las concentraciones recomendadas por CLSI y ASM.

Resultados: Las bacterias aisladas fueron: *Escherichia coli* (*E. coli*) enteropatógena (EPEC) 531, *Shigella* 120, *Salmonella* no-typhi 117, *Aeromonas* spp. 80, *E. coli* enterotoxigénica 54, *Yersinia enterocolitica* 20, *Campylobacter jejuni* 20, *Vibrio* spp. 20, *Pleisomonas shigelloides* 20 y *E. coli* enterohemorrágica (EHEC O:157) 18. La susceptibilidad global acumulada a RIF < 100, < 200, < 400, < 800 µg/ml fue del 70.6, el 90.8, el 99.3 y el 100%, respectivamente. La susceptibilidad global a cada antibiótico fue: AMP 32.2%, T-S 53.6%, NEO 54.1%, FUR 64.7%, CIP 67.3%, CLO 73%, FOS 81.3%. La susceptibilidad a RIF < 400 y < 800 µg/ml fue significativamente mayor que con los otros antimicrobianos ($p < 0.001$).

Conclusiones: La resistencia de las bacterias enteropatógenas a antimicrobianos utilizados en gastroenteritis es alta. La rifaximina fue activa contra el 99-100% de las bacterias en concentraciones alcanzables en el contenido intestinal con las dosis recomendadas.

© 2015 Asociación Mexicana de Gastroenterología. Publicado por Masson Doyma México S.A. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Acute diarrhea is an important health problem, mainly in the developing countries. Despite the decrease in mortality rates from this disease in the last decade in various countries, including Mexico¹⁻³, acute gastroenteritis continues to be a health problem due to its high morbidity. In Mexico, it is the second most common infectious disease, only preceded by respiratory diseases, with more than 5 million new cases per year⁴.

Despite the presence of certain clinical signs, it is difficult to determine the causal agent of acute diarrhea in a patient based solely on clinical findings. Acute gastroenteritis is often due to a viral infection, especially in children under 5 years of age¹, whereas bacterial infection is more habitual in older children and adults. The

most frequently detected enteropathogenic bacteria in patients with endemic gastroenteritis are *Escherichia coli* (*E. coli*) (EPEC, EIEC, EHEC, ETEC), *Campylobacter jejuni* (*C. jejuni*), *Shigella* spp, *Salmonella* spp, *Yersinia enterocolitica* (*Y. enterocolitica*)⁵⁻⁸, and less frequently, *Aeromonas* spp, *Vibrios* spp⁹⁻¹², and *Pleisomonas shigelloides* (*P. shigelloides*)¹³⁻¹⁵. The number of cases vary in relation to geographic region, patient age, and the season of the year in which the study was conducted.

The aims of therapeutic management of patients affected with gastroenteritis are to preserve life, relieve symptoms, prevent complications, cut the disease short, and prevent the spread of the pathogenic agents to the population. Oral rehydration is the standard treatment in acute gastroenteritis and antimicrobial agents are indicated in severe or prolonged cases, when shigellosis or cholera

are suspected, or when the pathogen is known, in order to prevent contagion¹⁶. However, in daily clinical practice, when the result of the stool culture is reported, the patient is already recovering or the treatment is delayed. Knowing the susceptibility of the bacteria causing a syndrome to the antimicrobial agents is important in 2 aspects: one is the early therapeutic approach, out of empirical necessity, and the other is the epidemiologic surveillance of bacterial resistance that is useful for taking measures to prevent it.

The empirical use of antimicrobials can be inefficient due to the emergence of bacterial resistance. Therefore the possible therapeutic regimens should be frequently updated, taking the pattern of regional resistance into account^{17–20}. Some clinical studies indicate that quinolones are superior to other antibiotics or to placebo in the empirical treatment of adult patients with diarrhea^{17,20–22}. Other antimicrobial drugs, such as ampicillin (AMP), trimethoprim-sulfamethoxazole (T-S), chloramphenicol (CHL), furazolidone (FUR), and non-absorbable antibiotics, such as neomycin and recently rifaximin (RIF), have been used in children, in whom quinolones are not indicated, and also in adults^{17,23–25}. Nevertheless, recent data on the bacterial susceptibility to these antimicrobial agents is not available in Mexico. RIF is a semisynthetic antibiotic derived from broad spectrum rifamycin S that inhibits the synthesis of bacterial RNA, is not absorbable when taken orally, and reaches a very high concentration in the intestinal lumen (~8,000 µg/g of feces). It has excellent bactericide activity on enteropathogenic microorganisms and does not cause important alterations in the gut microbiota²⁶.

The aim of this study was to investigate the susceptibility of the acute gastroenteritis-causing bacteria to rifaximin and the most widely used antibiotics in the treatment of gastrointestinal infections in Mexico.

Methods

Bacterial isolations

Bacterial strains obtained from the stool samples of 1,000 patients diagnosed with gastroenteritis or acute diarrhea were analyzed in 10 hospital laboratories in Mexico City that attend to hospitalized patients and outpatients. The strains were conserved and frozen before carrying out the biochemical identification, serology, and the antimicrobial susceptibility test in milk broth and soy broth with glycerol at -70°C ;²⁷ the initial primary isolations from the stool samples were carried out at each particular laboratory, and the following culture media were used: MacConkey Agar, Sorbitol-MacConkey Agar, *Salmonella Shigella* Agar, XLD Agar, *Campylobacter* Agar, *Yersinia* Agar, TCBS Agar, Brilliant Green Agar, and Tetrathionate Broth²⁸.

The isolates were biochemically identified in each laboratory using AutoScan 4, Walkaway (MicroScan), or Vitek 2 (Biomeriux) manual and automated processes and systems^{29–31} with an acceptance probability >95% in the identification; the typing and serologic identification in determined bacterial species, such as *E. coli*, *Salmonella*, and *Shigella*, were then performed using the Bio-Rad³², Phadebact³³, Oxoid³⁴, Sanofi-Pasteur³⁵, "O" Beli³⁶, and Probac³⁷ specific antisera and agglutination or coagglutination reagents. The detected serogroups and serotypes are shown in Table 1.

Antimicrobial agents and susceptibility tests

Antimicrobial susceptibility testing (AST) was done through the agar diffusion method (CHL, T-S, neomycin [NEO], FUR, fosfomicin [FOS], AMP, and ciprofloxacin [CIP]), and by

Table 1 Bacteria isolated from 1,000 patients diagnosed with acute gastroenteritis and their serotypes.

Bacteria	Number	Serotypes involved
Enteropathogenic <i>E. coli</i> group A, B, and C	531	O127:B8, O111:B4, O55:B5, O26:B6 and other serotypes O119:B4, O128:B12, O124:B17, O86:B7, O126:b16 and other serotypes O142:B86, O119:K90, O124:B17, O86:B7, O126:B16 and other serotypes that should correspond to: O128:K73, O44:K74, O18:K77, O20:K61, and O20:K84
Enterotoxigenic (LT) <i>E. coli</i>	54	Only thermolabile (TL) toxin-producing serotypes
Enterohemorrhagic <i>E. coli</i>	18	Only O:157 serotype
<i>Shigella dysenteriae</i>	54	All the serotypes that are agglutinable with the specific antiserum
<i>Shigella flexneri</i>	24	All the serotypes agglutinable with the specific antiserum
<i>Shigella boydii</i>	27	All the serotypes that are agglutinable with the specific antiserum
<i>Shigella sonnei</i>	15	All the serotypes that are agglutinable with the specific antiserum
<i>Salmonella</i> Group A	24	Most likely species: <i>S. paratyphi</i>
<i>Salmonella</i> Group B	21	Most likely species: <i>S. typhimurium</i>
<i>Salmonella</i> Group C1	24	Most likely species: <i>S. choleraesuis</i>
<i>Salmonella</i> Group C2	24	Most likely species: <i>S. newport</i>
<i>Salmonella</i> Group D	24	Most likely species: <i>S. enteritidis</i>
<i>Vibrio</i> spp.	20	–
<i>Yersinia enterocolitica</i>	20	–
<i>Campylobacter jejuni</i>	20	–
<i>Plesiomonas shigelloides</i>	20	–
<i>Aeromonas</i>	80	–

microdilution (RIF) following the recommendations of the Clinical & Laboratory Standards Institute³⁸ and the American Society for Microbiology³⁹. The concentrations of the antibiotics assayed and AST conditions were: AMP 10 µg/ml, T-S 1.25/23.75 µg/ml, CHL 30 µg/ml, CIP 5 µg/ml, FUR 100 µg/ml, NEO 30 µg/ml, FOS 50 µg/ml. RIF was tested at 100 µg/ml and the strains that were not susceptible at this concentration were successively exposed to concentrations of 200 µg/ml, 400 µg/ml, and 800 µg/ml. Susceptibility to RIF was considered at a minimal inhibitory concentration (MIC) of MIC₁₀₀, whereas it was MIC₉₀ for the other antibiotics.

Statistical analysis

Bacterial susceptibility to RIF was compared with the susceptibility to the other antimicrobial agents with the Z test and statistical significance was set at a $p < 0.05$. The Statistica 8.0 and Stata 11 statistical software were employed.

Results

The stool samples from 511 men and 489 women were analyzed. Sixty-five percent of the participants were above 20 years of age. The isolated bacteria were *E. coli* 603 (EPEC 531, ETEC 54, EHEC 18), *Shigella* 120 (*dysenteriae*, *flexneri*, *boydii*, *sonnei*), *Salmonella* 117 (*paratyphi*, *typhimurium*, *choleraesuis*, *newport*, *enteritidis*), *Vibrio* spp. 20, *Y. enterocolitica* 20, *C. jejuni* 20, *P. shigeloides* 20, and *Aeromonas* spp. 80.

Overall susceptibility of the bacteria to the different concentrations of RIF (MIC₁₀₀) are shown in Figure 1. The results of accumulated susceptibility patterns in all the strains assayed were at < 100 µg/ml: 70.6%; at < 200 µg/ml: 90.8%; at < 400 µg/ml: 99.3%; and at < 800 µg/ml: 100.0%. Table 2 shows the susceptibility of each group of bacteria to the different RIF concentrations. More than 99% of the strains of *Shigella*, *Salmonella*, *Yersinia*, *Campylobacter*, *Vibrio*, *Plesiomonas*, and *Aeromonas* were susceptible to concentrations < 100 or < 200 µg/ml, whereas 11-15% of the *E. coli* required < 400 or even < 800 µg/ml.

Overall susceptibility of all the bacterial species to each antibiotic was: RIF (< 400 µg/ml) 99.3% and (< 800 µg/ml)

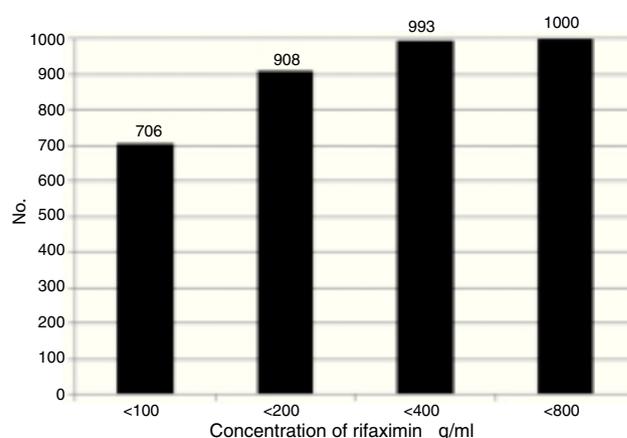


Figure 1 Overall accumulated susceptibility of the isolated bacteria to rifaximin. In the 1,000 strains, RIF was tested at a concentration of 100 µg/ml. The bacteria that were not susceptible at that concentration were successively exposed to concentrations of 200 µg/ml, 400 µg/ml, and 800 µg/ml. The accumulated susceptible strains were 706 (< 100), 908 (< 200), 993 (< 400), and 1,000 (< 800).

100%, FOS 81.3%, CHL 73.0%, CIP 67.3%, FUR 64.7%, NEO 54.1%, T-S 53.6%, and AMP 32.2%. Overall susceptibility to RIF with < 400 and < 800 µg/ml was superior ($p < 0.001$) to the overall susceptibility to each of the 8 drugs studied (fig. 2). This was also true in the majority of the cases in each bacterial group alone.

Table 3 shows the susceptibility of each bacterial group to the antibiotics tested.

As can be seen, a large number of strains show a very high proportion of bacterial resistance to NEO, T-S, and AMP, but *Yersinia* continues to be very susceptible to AMP (100%), *Vibrio* to T-S (90%), and *Campylobacter* to NEO.

Discussion

The prevalence and incidence of the bacteria that cause gastroenteritis worldwide vary according to the type of population studied, the geographic location of the study, the season of the year in which the microbiologic diagnosis is made, and other clinical and sociodemographic

Table 2 Susceptibility of the bacteria to different concentrations of rifaximin.

Bacteria	Number	< 100 µg/ml	< 200 µg/ml	< 400 µg/ml	< 800 µg/ml
EPEC	531	58.95	25.61	14.69	0.75
ETEC	54	79.63	7.41	7.41	5.56
EHEC	18	72.22	16.67	11.11	-
<i>Shigella</i>	120	90.00	10.00	-	-
<i>Salmonella</i>	117	67.52	31.62	0.85	-
<i>Yersinia</i>	20	80.00	20.00	-	-
<i>Campylobacter</i>	20	70.00	30.00	-	-
<i>Vibrio</i>	20	100.00	-	-	-
<i>Plesiomonas</i>	20	100.00	-	-	-
<i>Aeromonas</i>	80	100.00	-	-	-
All	1000	70.6	20.2	8.5	0.7

Results expressed in % of susceptible strains.

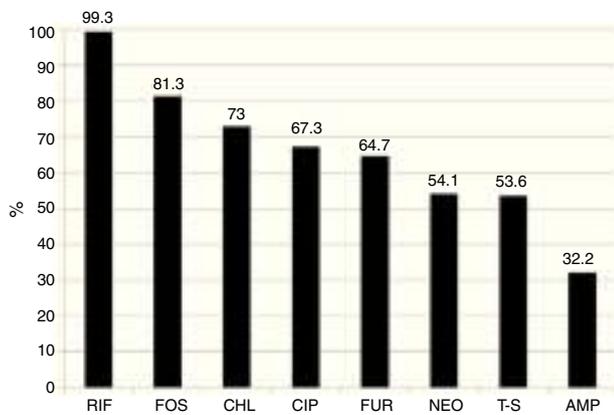


Figure 2 Overall susceptibility of the 1,000 isolated bacterial strains to the different antibiotics.

Rifaximin vs the other antimicrobials $p < 0.0001$.

AMP: ampicillin; CHL: chloramphenicol; CIP: ciprofloxacin; FOS: fosfomycin; FUR: furazolidone; NEO: neomycin; RIF: rifaximin ($< 400 \mu\text{g/ml}$); T-S: trimethoprim-sulfamethoxazole.

characteristics of the patients studied^{6,40–42}. Even though ours was not a prevalence study, the strains were collected throughout the entire year of 2013 and therefore the seasonal variations were not influential. The frequency of the strains received in the clinical analysis laboratories was reflected and the dominating ones were *E. coli*, *Shigella*, *Salmonella*, and *Aeromonas*. These data coincide with other reports in Mexico^{23,43,44}.

Standards for defining susceptibility or bacterial resistance have been published for the majority of antibiotics³⁸. Nevertheless, there are no clinical cut-off points for rifamycins against enteropathogens, although a value of $\leq 32 \mu\text{g/ml}$ of rifaximin was applied by some researchers in traveler's diarrhea^{45–47}. Bacterial susceptibility to an antibiotic implies that the isolations are inhibited by the usually achievable concentration of the antimicrobial agent at the infection site when the recommended dose is used. Fecal concentrations of rifaximin have been reported of up to $8,000 \mu\text{g/g}$ (mean $7,961 \mu\text{g/g}$) after 3 days of treatment at the recommended dose of 800mg/day , and there were still mean fecal concentrations of $3,266 \mu\text{g/g}$ 3 days after the

end of the therapy⁴⁸. In our study, we evaluated the susceptibility to rifaximin at 4 concentrations: < 100 , < 200 , < 400 , and $< 800 \mu\text{g/ml}$. The general susceptibility observed at these concentrations implies a certain bacterial resistance to rifaximin at concentrations of $< 100 \mu\text{g/ml}$ that was overcome at higher concentrations, but always much below the achievable values in the intestinal lumen. The cut-off points for considering bacteria to be resistant to rifaximin have been established at 32, 128, or even $256 \mu\text{g/ml}$ ^{46–49}, but, as observed in our study, the bacteria resistant to lower concentrations of rifaximin were not resistant if the concentration of the antibiotic was increased *in vitro*. Finally, 99.3% were susceptible at concentrations $< 400 \mu\text{g/ml}$ and 100% at concentrations $< 800 \mu\text{g/ml}$, which are 10–20 times lower than the mean concentration achieved *in vivo* with the recommended treatment.

The concentrations in the intestinal lumen, or fecal concentrations, of the other antimicrobial agents tested in the present study are not known, so the internationally recommended concentrations were used for the susceptibility analysis. In general, the bacteria were more susceptible to rifaximin than to the other antimicrobial agents tested in our analysis. Recent studies with enteropathogen isolates from travelers coming from Latin America and Asia also found a greater proportion of bacteria susceptible to rifaximin than to other antimicrobial agents. It was striking that the *C. jejuni* strains from Asia had a variable resistance to rifaximin, whereas those from Mexico and Guatemala were 100% susceptible^{46,47}. In our study, the *C. jejuni* strains were 100% susceptible at concentrations of < 100 – $200 \mu\text{g/ml}$. Several strains of *E. coli* required rifaximin concentrations greater than $100 \mu\text{g/ml}$, concurring with other observations on bacteria coming from Mexico⁵⁰.

Rifaximin has a broad spectrum of susceptibility that includes anaerobic bacteria^{26,49}. Bacterial resistance is very low and is not easily induced⁵¹. In addition, Ouyang–Latimer et al. compared the susceptibility of enteropathogenic bacteria obtained in 2006–2008 with those obtained 10 years earlier and found no increase in the MIC₉₀ levels for rifaximin in any of the organisms analyzed⁴⁶. In fact, the normal intestinal bacteria that became resistant to $100 \mu\text{g/ml}$ of rifaximin after 5 days of treatment spontaneously disappeared from the stools within a few weeks⁵².

Table 3 Susceptibility of each group of bacteria to the antimicrobials tested.

Group of bacteria	FOS	CHL	CIP	FUR	NEO	T-S	AMP
EPEC	7.1	66.6	51.9	66.4	48.5	49.1	27.6
ETEC	75.9	88.8	50.0	77.7	53.7	50.0	29.6
EHEC	55.5	83.3	100	50.0	61.1	16.6	16.6
<i>Shigella</i>	68.3	82.5	97.5	47.5	46.6	27.5	48.3
<i>Salmonella</i>	77.8	64.1	89.7	64.1	52.1	74.4	50.4
<i>Yersinia</i>	90.0	80.0	90.0	80.0	30.0	2.5	100
<i>Campylobacter</i>	80.0	95.0	30.0	100	100	30.0	45.0
<i>Vibrio</i>	90.0	80.0	100	90.0	70.0	90.0	55.0
<i>Plesiomonas</i>	100	95.0	80.0	85.0	90.0	90.0	20.0
<i>Aeromonas</i>	67.5	86.0	87.5	50.0	80.0	78.7	20.0
All	81.3	73.0	67.3	64–7	54.1	53.6	32.2

Results expressed in % of susceptible strains.

The low proportion of bacteria that are susceptible to AMP, T-S, neomycin, FUR, and CIP, from 32.2 to 67.3% in that order, coincides with studies conducted in other regions of the world⁵³⁻⁵⁶ and can be explained by the induction of bacterial resistance due to the widespread use or abuse of those antimicrobial agents. In this respect, it is striking that CHL, which conserves an acceptable general susceptibility of 73%, has been restricted, after its widespread use in Mexico, to treating typhoid fever for the last 30 years. And comparatively, in our study we introduced the test of susceptibility to FOS. This antibiotic is used very little in Mexico, and mainly in urinary infections. The general susceptibility of FOS was greater than that of the other antimicrobial agents, just below rifaximin.

Neomycin is a poorly absorbed antibiotic that is widely used in Mexico and does not require a prescription. According to our data, it is clear that AMP, T-S, neomycin, and FUR would not be good therapeutic options. RIF has been successfully used for treating acute infectious diarrhea in children and adults, as well as for traveler's diarrhea, with excellent tolerance⁵⁷⁻⁶⁰. RIF would appear to be the best option, given that it has a high therapeutic index, combining high efficaciousness with a low frequency of adverse effects²⁶. Adverse effects are frequently attributed to other antibiotics, especially to beta-lactams and T-S mainly due to allergies, and the sulfonamides and CIP are known to be neurotoxic⁶¹.

Treatment with antimicrobial agents is a valuable tool in the control of several gastrointestinal infections, given that the length of time and intensity of the disease is reduced, potentially severe complications are prevented, and disease transmission is decreased. Unfortunately, for several decades, enteropathogenic bacteria strains have been selected that are resistant to the commonly used antimicrobial agents and to those that once were considered first choice⁶². The selection and dissemination of antimicrobial resistance among the different bacterial species that cause gastroenteritis is a growing health problem that complicates the therapeutic management of the severe cases. Studies conducted in many parts of the world have revealed an important increase in antimicrobial-resistant bacteria in many infectious diseases, including gastroenteritis^{63,64}. To reduce the appearance of antimicrobial-resistant bacterial strains, it would seem prudent to avoid the indiscriminate use of antibiotics in cattle, periodically determine local bacterial resistance patterns, reduce the easy access to antimicrobial drugs (for example: prevent self-medication with antimicrobials), and strengthen the medical education on this topic.

Study limitations. The stool cultures were obtained from 10 laboratories that sent positive stool cultures with enteropathogen bacteria from patients diagnosed with acute gastroenteritis or acute diarrhea. Therefore, it was not possible to obtain the clinical data, such as the length of time of symptom progression, stool characteristics, the symptoms accompanying the diarrhea, and the intensity or severity of the clinical symptoms that would enable their correlation with the genus and species of the bacterial isolates. The stool cultures were from Mexico City and do not necessarily reflect those from the rest of the country.

Ethical responsibilities

Protection of persons and animals. The authors declare that no experiments were performed on humans or animals for this study.

Data confidentiality. The authors declare that no patient data appear in this article.

Right to privacy and informed consent. The authors declare that no patient data appear in this article.

Financial disclosure

This study was financially supported by Alfa Wassermann S.A. de C.V.

Conflict of interest

Alberto Frati is presently the medical director of Alfa Wassermann S.A. de C.V., which produces rifaximin. He participated in the planning of the project and manuscript revision, but did not intervene in any phase of the study, the enumeration of the results, or the statistical analysis. The other authors declare that there is no conflict of interest.

References

1. Velázquez FR, García-Lozano H, Rodríguez E, et al. Diarrhea morbidity and mortality in Mexican children: Impact of rotavirus disease. *Pediatr Infect Dis J*. 2004;23 10 Supl:S149-55.
2. Farthing M, Lindberg G, Dite P, et al. Acute diarrhea. World Gastroenterology Organization. 2008. WGO Practice Guidelines. p. 1-31.
3. Esparza Aguilar M, Bautista- Márquez A, González-Andrade MD, et al. Mortalidad por enfermedad diarreica en menores, antes y después de la introducción de la vacuna contra el rotavirus. *Salud Pub Mex*. 2009;51:285-90.
4. SUIVE/DGE/SALUD/información epidemiológica de morbilidad. Anuario 2011.
5. Gallegos M, Morales A, Álvarez G, et al. Caracterización de aislados de *Escherichia coli* O157: H7 en canales de bovinos y porcinos mediante PCR. Marzo 2009. *Rev Científica (Maracaibo)*. 2009;10:1-15.
6. Sánchez MLG, González LL. *Yersinia enterocolitica*: prevalencia en niños con diarrea atendidos en el Hospital Municipal de Córdoba durante 2010-2011 [accessed 10 Aug 2014]. Available from: <http://www.cobico.com.ar/wp-content/archivos/2013/02/PUBLICACION-DRA-SANCHEZ.pdf>.
7. Mirón D, Schotnick I, Yardeni D, et al. Surgical complications of Shigellosis in children. *Pediatr Infect Dis J*. 2000;19:898-900.
8. Campos G, Alarcón T, Domingo D, et al. Sensibilidad de *Campylobacter jejuni* a ocho antibióticos en cepas aisladas de muestras clínicas de niños. *Rev Esp Quimioterap*. 2003;16:216-20.
9. Cabrera RL, Bravo FL, Ramírez AMM, et al. Susceptibilidad a los antimicrobianos y factores de virulencia en cepas de *Vibrio cholerae* no-01 aisladas de pacientes con enfermedad diarreica aguda. *Rev Biomed*. 2008;19:138-44.
10. Evaluación de riesgos de *Vibrio* spp. en pescados y mariscos. Ginebra: WHO/FAO 2004 [accessed 20 Aug 2014]. Available from: <http://www.fao.org/docrep/008/y8145s/y8145s08.htm#TopOfPage>

11. Muñoz D, Castañeda de F, Grau de M, et al. Prevalencia y susceptibilidad a los antibióticos de cepas móviles de *Aeromonas* aisladas del ostión de mangle (*Crassostrea rhizophorae*). Rev Científica. 2012;22:565–73.
12. Bravo L, Fernández A, González D, et al. Caracterización fenotípica de cepas de *Aeromonas* aisladas de pacientes con enfermedad diarreica aguda en Cuba. Rev Chil Infect. 2011;28:159–65.
13. Cravioto A, Tello A, Navarro A, et al. Association of *Escherichia coli* Hep-2-adherence patterns with the type and duration of diarrhea. Lancet. 1991;337:262–4.
14. Nataro JP, Kaper JB. Diarrheagenic *Escherichia coli*. Clin Microbiol Rev. 1998;11:142–201.
15. Torrey S, Fleisher G, Jaffe D, et al. Incidence of bacteremia in infants with *Salmonella enteritidis*. J Pediatr. 1986;108:718–21.
16. Farthing M, Lindberg G, Dite P, et al. World Gastroenterology Organization practice guideline: Acute diarrhea. WGO Practice Guidelines. 2008:1–29.
17. Solórzano F, Miranda N. Resistencia de bacterias respiratorias y entéricas a los antibióticos. Salud Pub México. 1998;40:510–6.
18. Dryden MS, Gabb RJ, Wright SK. Empirical treatment of severe acute community-acquired gastroenteritis with ciprofloxacin. Clin Infect Dis. 1996;22:1019–25.
19. Goodman LJ, Trenholme GM, Kaplan RL, et al. Empiric antimicrobial therapy of domestically acquired acute diarrhea in urban adults. Arch Intern Med. 1990;150:541–6.
20. Prado V, Pidal P, Arellano C, et al. Multirresistencia antimicrobiana en cepas de *Shigella* spp. en una comuna semirural del área norte de Santiago. Rev Med Chil. 1998;126:1464–71.
21. Sánchez C, García-Restoy E, Garau J, et al. Ciprofloxacin and trimetropin-sulfametoxazol versus placebo in acute uncomplicated *Salmonella enteritidis*: A double-blind trial. J Infect Dis. 1998;168:1304–7.
22. Dupont L, Dong Jiang Z, Ericsson C, et al. Rifaximin versus ciprofloxacin for the treatment of traveler's diarrhea: A randomized, double-blind clinical Trial. Clin Infect Dis. 2001;33:1807–15.
23. Patrick Basu P, Dinami R, Rayapudi K. Rifaximin therapy for metronidazole-unresponsive *Clostridium difficile* infection: A prospective pilot trial. Ther Adv Gastroenterol. 2010;3:221–5.
24. Gobernado M, Ponce J. Rifaximina. Rev Esp Quimioterap. 2004;17:141–53.
25. Pimentel M, Lembo A, Chey W, et al. Rifaximin therapy for patients with Irritable Bowel syndrome without constipation. N Engl J Med. 2011;364:22–32.
26. Scarpignato C, Pelosini I. Experimental and clinical pharmacology of rifaximin, a gastrointestinal selective antibiotic. Digestion. 2006;73 Suppl 1:13–27.
27. Sánchez-Leal LC, Corrales Ramírez LC. Evaluación de la congelación para la conservación de especies bacterianas. Nova-Pub Cient. 2005;3:1–116.
28. Zimbro MJ, Power DA, Miller SM, et al., eds. Difco & BBL Manual. Manual of microbiological culture media. 2nd ed. Sparks: BD Diagnostics-Diagnostic Systems; 2009.
29. Instructivo Panel Gram Positivo. MicroScan. Siemens Healthcare Diagnostic, México, 2013.
30. Instructivo Panel ID-GNB. Vitek 2 Biomeriux Inc. México, 2013.
31. Instructivo Panel Gram Negativo. MicroScan. Siemens Healthcare Diagnostics. México, 2013.
32. Instructivos Bio-rad Laboratories. Antisueros bacterianos. México, 2013.
33. Instructivos Phadebact Monoclonal GC Test. Suecia, 2012.
34. Instructivos Oxoid Limited. Dryspot *E. coli* O 157. United Kingdom, 2013.
35. Instructivos Sanofi-Pasteur. Antisueros bacterianos. 2012. México. France.
36. Instructivos Laboratorios «O» Beli. Antisueros bacterianos. 2012. México DF.
37. Instructivos Probac do Brazil. Antisueros bacterianos. 2012. Sao Paulo, Brazil.
38. Clinical Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Twenty-second informational supplement. M100-S22 2012; 32 (3).
39. Koneman EW, Allen SD, Janda WM, et al. Diagnostic Microbiology. 5th ed. Lippincott: Philadelphia; 2000. p. 131–6.
40. Bravo L, Fernández A, González D, et al. Caracterización fenotípica de cepas de *Aeromonas* aisladas de pacientes con enfermedad diarreica aguda en Cuba. Rev Chil Infect. 2011;28:159–65.
41. Njuguna HL, Cosmas L, Williamson J, et al. Use of population-based surveillance to define the high incidence of shigellosis in an urban slum in Nairobi, Kenya. PLoS ONE. 2014;9:e105031.
42. Amisano G, Fornasero S, Migliaretti G, et al. Diarrheagenic *Escherichia coli* in acute gastroenteritis in infants in North-West Italy. New Microbiol. 2011;34:45–51.
43. Duncan B, Fulginiti VA, Sieber OF, et al. Shigella sepsis. Am J Dis Child. 1981;135:151–4.
44. Rodríguez Ángeles G. Principales características y diagnóstico de los grupos patógenos de *Escherichia coli*. Salud Pub Mex. 2002;44:464–75.
45. Gomi H, Jiang ZD, Aldachi JA, et al. *In vitro* antimicrobial susceptibility testing of bacterial enteropathogens causing traveler's diarrhea in four geographic regions. Antimicrob Agents Chemother. 2001;45:212–6.
46. Ouyang-Latimer J, Jafri S, VanTassel A, et al. In vitro Antimicrobial susceptibility of bacterial enteropathogens isolated from international travelers to Mexico, Guatemala, and India from 2006 to 2008. Antimicrob Agents Chemother. 2011;55:874–8.
47. Hopkins KL, Mushtaq S, Richardson JF, et al. *In vitro* activity of rifaximin against clinical isolates of *Escherichia coli* and other enteropathogenic bacteria isolated from travelers returning to the UK. Int J Antimicrob Agents. 2014;43:431–7.
48. Jiang ZD, Ke S, Palazzini E, et al. In vitro activity and fecal concentration of rifaximin after oral administration. Antimicrob Agents Chemother. 2000;44:2205–6.
49. Wang FD, Liao CH, Lin YT, et al. Trends in the susceptibility of commonly encountered clinically significant anaerobes and susceptibilities of blood isolates of anaerobes to 16 antimicrobial agents, including fidaxomicin and rifaximin, 2008–2012, northern Taiwan. Eur J Clin Microbiol Infect Dis. 2014;33:2041–52.
50. Jiang ZD, DuPont HL. Rifaximin: *in vitro* and *in vivo* antibacterial activity —A review. Chemotherapy. 2005;51 Suppl 1:67–72.
51. DuPont HL, Jiang ZD. Influence of rifaximin treatment on the susceptibility of intestinal Gram-negative flora and enterococci. Clin Microbiol Infect. 2004;10:1009–11.
52. De Leo C, Eftimiadi C, Schito GC. Rapid disappearance from the intestinal tract of bacteria resistant to rifaximin. Drugs Exptl Clin Res. 1986;12:979–81.
53. Barati S, Boniadian M, Habibian R, et al. Antibiotics resistance of enterotoxigenic and enteroaggregative *Escherichia coli* isolated from gastroenteritis cases. Asian J Biomed Pharm Sci. 2012;2:54–8.
54. Yang H, Chen G, Zhu Y, et al. Surveillance of Antimicrobial susceptibility patterns among *Shigella* species isolated in China during the 7-year period of 2005–2011. Ann Lab Med. 2013;33:111–5.
55. Li Y, Xie X, Xu X, et al. Non typhoidal *Salmonella* infection in children with acute gastroenteritis: Prevalence, serotypes and antimicrobial resistance in Shanghai, China. Foodborne Pathog Dis. 2014;11:200–6.
56. Marcoleta A, Toro C, Prado V, et al. Patrones de susceptibilidad antimicrobiana de cepas de *Shigella sonnei* aisladas durante tres periodos diferentes en la región metropolitana. Chile. Rev Chil Infect. 2013;30:616–21.
57. Macías Parra M, González Saldaña N, Ramírez Sandoval P, et al. Eficacia y seguridad de rifaximina en el tratamiento del episodio

- diarréico agudo en niños de seis meses a cinco años. *Rev Enf Inf Pediat*. 2002;16:23–8.
58. Della Marchina M, Renzi G, Palazzini E. Infectious diarrhea in the aged: Controlled clinical trial of rifaximin. *Chemiother*. 1988;7:336–40.
 59. Taylor DN, Bourgeois AL, Ericsson CD, et al. A randomized, double-blind, multicenter study of rifaximin compared with placebo and with ciprofloxacin in the treatment of travelers' diarrhea. *Am J Trop Med Hyg*. 2006;74:1060–6.
 60. DuPont HL, Jiang ZD, Belkin-Gerson J, et al. Treatment of travelers' diarrhea: A randomized trial comparing rifaximin, rifaximin plus loperamide and loperamide alone. *Clin Gastroenterol Hepatol*. 2007;5:451–6.
 61. Shehab N, Patel PR, Srinivasan A, et al. Emergency department visits for antibiotic-associated adverse events. *Clin Infect Dis*. 2008;47:735–43.
 62. Vrints M, Mairiaux E, van Meervenne E, et al. Surveillance of antibiotic susceptibility patterns among *Shigella sonnei* strains isolated in Belgium during the 18-year period 1990 to 2007. *J Clin Microbiol*. 2009;47:1379–85.
 63. Fullá N, Prado V, Durán C, et al. Surveillance for antimicrobial resistance profiles among *Shigella* species isolated from a semirural community in the northern administrative area of Santiago, Chile. *Am J Trop Med Hyg*. 2005;72:851–4.
 64. Sack RB, Rahman M, Yunus M, et al. Antimicrobial resistance in organisms causing diarrheal disease. *Clin Infect Dis*. 1997;24 Suppl 1:S102–5.