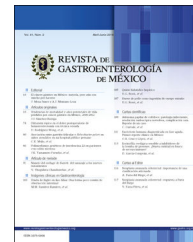




# REVISTA DE GASTROENTEROLOGÍA DE MÉXICO

[www.elsevier.es/rgmx](http://www.elsevier.es/rgmx)



## ORIGINAL ARTICLE

# Evaluation of chronic diarrhea in patients newly diagnosed with HIV infection through the FilmArray® gastrointestinal panel

R. Montalvo-Otivo<sup>a,\*</sup>, P. Vilcapoma<sup>b</sup>, A. Murillo<sup>c</sup>, C. Mathey<sup>b</sup>, A. Olivera<sup>d</sup>, G. Veliz<sup>d</sup>, D. Estrella<sup>e</sup>

<sup>a</sup> Facultad de Medicina Humana, Universidad Cesar Vallejo, Peru

<sup>b</sup> Facultad de Medicina Humana, Universidad Continental, Huancayo, Peru

<sup>c</sup> Servicio de Enfermedades Infecciosas, Hospital Daniel Alcides Carrión, Huancayo, Peru

<sup>d</sup> Facultad de Medicina Humana, Universidad Peruana Los Andes, Huancayo, Peru

<sup>e</sup> Servicio de Microbiología, Hospital Daniel Alcides Carrión, Huancayo, Peru

Received 25 May 2022; accepted 16 November 2022

### KEYWORDS

HIV/AIDS;  
Chronic diarrhea;  
Automated nested  
multiplex polymerase  
chain reaction  
system;  
Diagnoses

### Abstract

**Introduction:** The treatment and diagnosis of chronic diarrhea in the immunocompromised patient depends on the ability to rapidly detect the etiologic agents.

**Aims:** Our aim was to evaluate the results of the FilmArray® gastrointestinal panel in patients newly diagnosed with HIV infection that presented with chronic diarrhea.

**Material and methods:** Utilizing nonprobability consecutive convenience sampling, 24 patients were included that underwent molecular testing for the simultaneous detection of 22 pathogens.

**Results:** In 24 HIV-infected patients with chronic diarrhea, enteropathogen bacteria were detected in 69% of the cases, parasites in 18%, and viruses in 13%. Enteropathogenic *Escherichia coli* and enteroaggregative *Escherichia coli* were the main bacteria identified, *Giardia lamblia* was found in 25%, and norovirus was the most frequent viral agent. The median number of infectious agents per patient was three (range of 0 to 7). The biologic agents not identified through the FilmArray® method were tuberculosis and fungi.

**Conclusions:** Several infectious agents were simultaneously detected through the FilmArray® gastrointestinal panel in patients with HIV infection and chronic diarrhea.

© 2023 Asociación Mexicana de Gastroenterología. Published by Masson Doyma México S.A. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

\* Corresponding author. Calle Anís n.º 210, El Tambo, Huancayo, Peru. Tel.: +51 992406768.  
E-mail address: [otivo3@hotmail.com](mailto:otivo3@hotmail.com) (R. Montalvo-Otivo).

**PALABRAS CLAVE**

VIH/Sida;  
Diarrea crónica;  
Automated nested  
multiplex polymerase  
chain reaction  
system;  
Diagnósticos

## Evaluación de la diarrea crónica en pacientes con reciente diagnóstico de infección por VIH mediante el panel gastrointestinal FilmArray®

**Resumen**

**Introducción:** El tratamiento y el pronóstico de la diarrea crónica en el paciente inmunosuprimido dependen de la capacidad para detectar rápidamente los agentes etiológicos.

**Objetivos:** Evaluar los resultados del panel gastrointestinal (GI) FilmArray® en pacientes con diarrea crónica e infección por VIH de reciente diagnóstico.

**Material y métodos:** Muestreo no probabilístico a conveniencia de manera secuencial. Se incluyeron 24 pacientes a quienes se realizó el estudio molecular para la detección simultánea de 22 patógenos.

**Resultados:** En 24 pacientes infectados por el VIH con diarrea crónica se detectaron bacterias enteropatógenas en el 69% de los casos, parásitos en el 18% y virus en el 13%. *Escherichia coli* enteropatógena y *Escherichia coli* enteroagregativa fueron las principales bacterias identificadas; *Norovirus* fue el agente viral más frecuente encontrado (33%), seguido de *Giardia lamblia* (25%). La mediana del número de agentes infecciosos por paciente fue 3 (rango de 0 a 7). Los agentes biológicos no identificados por este método fueron tuberculosis y hongos.

**Conclusiones:** Utilizando el panel GI FilmArray® se detectaron simultáneamente varios agentes infecciosos en pacientes con diarrea crónica e infección por VIH.

© 2023 Asociación Mexicana de Gastroenterología. Publicado por Masson Doyma México S.A. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Introduction and aim**

Diarrhea is a common morbidity and is responsible for over one million annual deaths worldwide<sup>1</sup>. If symptoms persist for more than four weeks, the condition is known as chronic diarrhea, which is a problem that affects up to 5% of the population at any given time<sup>2</sup>. It can be fatal in immunocompromised patients and occurs at a rate at least 10 times higher in individuals infected with human immunodeficiency virus (HIV), compared with the general population<sup>3</sup>. Diarrhea can affect patients with HIV at all stages of the disease and up to 60% of said patients report having symptoms of diarrhea<sup>4</sup>. Despite antiretroviral therapy (ART), HIV-associated diarrhea continues to be of multifactorial etiology that includes infectious processes, neoplasia, and enteropathy<sup>5</sup>, producing quality of life alterations, antiretroviral treatment adherence failure, weight loss, and malnutrition<sup>6,7</sup>.

The search for the etiologic agent of chronic diarrhea in the patient with HIV includes the performance of a microbiologic stool analysis (mainly stool culture, different methods for detecting intestinal parasites, and bacilloscopy), molecular methods (polymerase chain reaction [PCR]), clinical evaluation (considering the CD4 T lymphocyte count and HIV viral load determination), abdominal tomography, and colonoscopy<sup>8</sup>. Multiple infectious agents have been observed to cause chronic diarrhea and the complete clinical picture often cannot be explained by the identification of a single etiologic agent, with liquid stools persisting despite the treatment regimen, resulting in the continued performance of diagnostic tests.

Inopportune etiologic agent identification can cause significant morbidity and be responsible for approximately 80% of acquired immune deficiency syndrome (AIDS)-related

deaths<sup>9</sup>. This type of diarrhea is difficult to manage and the etiologic agent is often not identified. In addition, physical examination and microbiologic test results may not be sufficient for directing therapy in some patients, at times leading the physician to resort to using antidiarrheals to improve the patient's quality of life<sup>10</sup>.

*Salmonella* spp., *Shigella* spp., *Campylobacter* spp., and *Mycobacterium tuberculosis* are among the most frequent bacterial agents; *Cystoisospora belli*, *Cryptosporidium parvum*, *Cyclospora cayetanensis*, and *Microsporidia* are among the parasitic agents; and cytomegalovirus or herpes simplex virus type 1 and type 2 are among the viral agents. Lastly, invasive fungi, such as *Histoplasma capsulatum* and *Paracoccidioides brasiliensis* have been reported. Etiologic agent identification is generally difficult due to multiple factors, such as self-medication before seeking medical care, the poor diagnostic methods available at many laboratories, and the special analysis technique each microbial agent requires<sup>11</sup>.

One of the molecular methods available for simultaneously identifying several infectious agents is the FilmArray® Gastrointestinal (GI) Panel (MEP, BioFire Diagnostics/Biomérieux, Salt Lake City, UT, USA). It is a PCR-based technique that is useful for the rapid simultaneous detection of 22 pathogens (13 bacteria, 5 viruses, and 4 parasites) from liquid stool samples<sup>12</sup>. However, in immunocompromised patients, its true performance, compared with conventional diagnostic methods, is still unknown, especially in patients with HIV.

The aim of the present study was to evaluate the infectious agents that cause chronic diarrhea in patients with HIV infection, utilizing the FilmArray® GI Panel. Ours is the first study to utilize this molecular method for etiologic agent

identification in HIV patients presenting with chronic diarrhea.

## Materials and methods

### Study design

An observational, analytic, cross-sectional study was conducted on patients that were seen at the Tropical and Infectious Diseases Service of the *Hospital Daniel Alcides Carrión (HDAC)* in Huancayo, Peru, within the time frame of January to December 2021. The inclusion criteria were age above 18 years, HIV infection diagnosis, watery stools for more than 4 weeks, a CD4 T lymphocyte count, and viral load for HIV. Liquid stool samples were collected to identify the etiologic agent of the chronic diarrhea.

The variables of clinical data, time of disease, treatment regimen, CD4 cell count, viral load, and ART were collected. All participants gave their informed consent.

### Statistical analysis

Nonrandomized consecutive sampling was carried out until the sample size, determined using the Stata version 14 program, was reached. To calculate the sample size, the main limiting factor was assumed to be the number of positive cases, given that its precision would depend on the identification of the infectious agent and clinical improvement after receiving the specific treatment. With a 10% margin of error, an 80% power, and a supposed minimally acceptable sensitivity of 80% for identifying the etiologic agent<sup>13,14</sup>, we calculated that a minimum of 24 patients with HIV infection and chronic diarrhea should be enrolled. The double data entry table was carried out utilizing the Excel program. The number and percentage of patients for the categorical variables and the median (interquartile range [IQR]) for the continuous variables were shown.

### Procedure

After identifying the patients that met the inclusion criteria, they were invited to participate in the study. After providing their statements of informed consent, the patients were instructed to collect their stool in a screw cap jar for the stool culture samples. The samples were received at the laboratory and immediately processed through the FilmArray® GI Panel. Pasty or solid stool samples were rejected because the Cary-Blair medium for diluting the samples and enabling the correct processing of the panel was not available. The samples were not stored for later testing due to the fact that time and temperature could affect future procedures. Thus, all processes were carried out as soon as the samples arrived at the laboratory.

In patients that continued to present with chronic diarrhea despite receiving the treatment regimen against the infectious agents identified by the FilmArray® GI Panel, or when the panel results were negative, colonoscopy with biopsy taken at the affected area, performed by a gastroenterologist, was ordered. The modified Ziehl-Neelsen, PAS, and Grocott Gomori stains were prioritized, in addition

to conventional stains, for the histopathologic study of the samples.

### FilmArray® Gastrointestinal Panel

The FilmArray® GI Panel is an *in vitro* multiplexed diagnostic test with a closed system that contains all the reagents needed for inverse transcription, PCR, and the detection of nucleic acid of different gastrointestinal pathogens in a single stool sample. The system is approved by the FDA in the United States and by the European Union.

Said gastrointestinal panel identifies 22 pathogens that include: bacteria (*Campylobacter jejuni*, *Campylobacter coli*, *Campylobacter upsaliensis*, *Clostridium difficile* (A/B toxins), *Plesiomonas shigelloides*, *Salmonella* spp., *Yersinia enterocolitica*, *Vibrio parahaemolyticus*, *Vibrio vulnificus*, *Vibrio cholerae*, *Shigella* spp. and pathotypes of diarrheagenic *Escherichia coli* (*E. coli*), such as enteroaggregative *E. coli* (EAEC), enteropathogenic *E. coli* (EPEC), enterotoxigenic *E. coli* (ETEC) LT/ST, Shiga toxin-producing *E. coli* (STEC) stx1/stx2, *E. coli* O157, and enteroinvasive *E. coli* (EIEC); parasites (*Cryptosporidium*, *Cyclospora cayetanensis*, *Entamoeba histolytica*, *Giardia lamblia*), and viruses (adenovirus F 40/41, astrovirus, norovirus GI/GII, rotavirus A, sapovirus (I, II, IV, and V).

Samples were processed according to the manufacturer's instructions (The BioFire® FilmArray® Gastrointestinal Panel). The processing time of each sample was about an hour, and the results were immediately reported to the treating physician.

A result was considered positive or negative when the final software report was obtained.

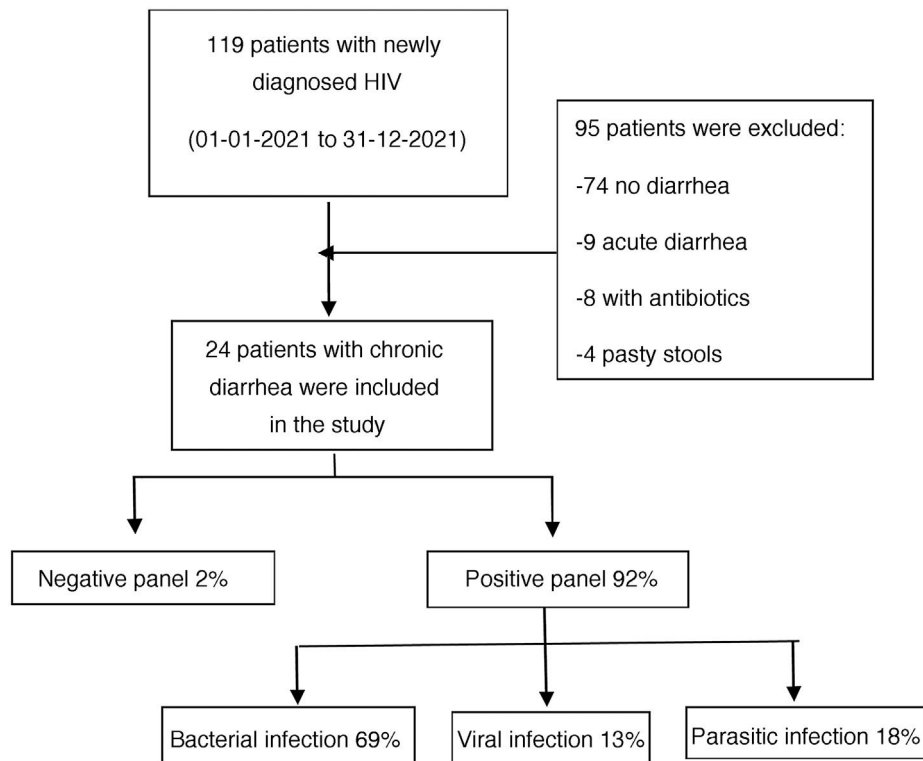
### Ethical considerations

This work was approved by the ethics committee of the *Hospital Daniel Alcides Carrión (HDAC)* in Huancayo, Peru, with authorization number 018-2021, respecting patient identification through codes and prior informed consent.

## Results

Within the time frame of January to December 2021, 119 patients newly diagnosed with HIV were evaluated at the Infectious Diseases Service of the *HDAC*. Twenty-four of those patients met the inclusion criteria and were enrolled in the study (Fig. 1). All the patients fit the definition of newly diagnosed HIV infection and chronic diarrhea. Eighteen of the participants were males and the median patient age was 37 years, with an IQR of 26 to 75 years of age.

The median CD4 T lymphocyte count was 35 cells per mm<sup>3</sup>, with an IQR of 5 to 231 cells per mm<sup>3</sup>; the median viral load was 39,400 copies/ml, with an IQR of 2,510 to 120,000 copies/ml; 8 (30%) patients had started ART in the past month, all of whom took one tablet daily of the dolutegravir 50 mg/lamivudine 300 mg/tenofovir 300 mg coformulation; 16 (70%) patients were diagnosed at the Infectious Diseases



**Figure 1** Description of the study population, within the January to December 2021 study period.

Service and were not undergoing ART at the time of their enrollment in the study (Table 1).

According to the etiologic agent grouped by type of infectious microorganism, in the positive samples, bacteria were responsible for the chronic diarrhea in 69% of the patients, parasites were identified in 18%, and viruses in 13%.

In the 24 watery stool samples that were positive, *E. coli* was the most frequently identified bacterial agent in chronic diarrhea and the most frequent of those diarrheagenic pathotypes were EPEC, EAEC, and ETEC, at values of 51%, 40%, and 29%, respectively. The second most frequently identified bacterium was *Campylobacter jejuni*, found in 26% of the samples. Norovirus was the most prevalent of the viral agents, detected in 33% of the samples, and sapovirus was identified in 3%.

*Giardia lamblia* was the most frequent parasite, found in 25% of the samples, followed by *Cryptosporidium* spp., identified in 22% of the patients with chronic diarrhea.

Three was the median number of infectious agents found in a single patient with HIV, with a range of 0 to 7 infectious microorganisms per patient. In the 24 patients analyzed, 2 (8%) had negative samples and 22 (92%) had positive samples. Table 2 provides a detailed summary of the performance of the FilmArray® GI Panel for each of the 24 patients.

Two of the 24 patients did not respond to the treatment regimen against the identified infectious agent and a pathogen could not be identified in another two patients, and so those four patients underwent colonoscopy. *M. tuberculosis* was found in two of them, *Histoplasma* spp. in one, and mucormycosis in one (Table 2).

## Discussion

Concurrent with previous studies, diarrhea is the most frequent morbidity in patients with HIV infection, and without ART, there is up to an 80% possibility of developing chronic diarrhea. The most frequent causes of such enteric infections are mainly bacterial in origin<sup>15</sup>. In one study, parasites were the main cause of chronic diarrhea<sup>16</sup>, whereas in another, tuberculosis was the primary cause<sup>17</sup>, each with a different epidemiologic history.

Despite all the existing methods, there is still a group of patients in whom the pathogenic agent that causes chronic diarrhea cannot be identified, resulting in malnutrition and physical deterioration, known as HIV enteropathy<sup>18</sup>, and becoming a public health problem because of the inadequate use of numerous antibiotics and increased antimicrobial resistance. Starting ART early is known to decrease those infections, but it is also related to the ability to detect the etiologic agents associated with chronic diarrhea.

The FilmArray® GI Panel has been evaluated for detecting the infectious agent in enteritis in immunocompromised patients and has 100% sensitivity and 97% specificity, when compared with conventional cultures<sup>19</sup>. It also has a capacity of up to 40% for detecting agents that are not commonly studied<sup>20</sup>.

However, there are few reports on immunocompromised patients; in one case of multiple myeloma with chronic diarrhea, the agent identified using the FilmArray® GI Panel was *E. coli*<sup>21</sup>. In our study, the agents most frequently identified were bacteria, similar to that reported in a study from the United States in patients with chronic diarrhea,

**Table 1** General characteristics of the patients with HIV infection and chronic diarrhea.

Characteristic	Results (n = 24)
Age years, median (IQR)	37 (26-75)
Male sex	18 (75%)
CD4 (cell/mm <sup>3</sup> ), median (IQR)	35 (5-231)
Viral load, median (IQR) (copies/ml)	39,400 (2,510-120,000)
Moderate-to-severe diarrhea	7 (25%)
Undergoing antiretroviral treatment	8 (30%)
Newly diagnosed with HIV	16 (70%)
Negative result	2 (8%)
Positive result	22 (92%)
Number of infectious agents identified per patient, median (IQR)	3 (0-7)
Number of infectious agents according to CD4 T lymphocyte count:	
≤100; median (IQR)	4 (1-6)
>100; median (IQR)	2 (2-5)
Identification by etiologic group:	
Bacteria	52 (69%)
Parasites	13 (18%)
Viruses	10 (13%)
Specific etiologic agent:	
Enteropathogenic <i>E. coli</i>	13 (51%)
Enteropathogenic <i>E. coli</i>	10 (40%)
Norovirus	9 (33%)
Enterotoxigenic <i>E. coli</i>	7 (29%)
<i>Campylobacter jejuni</i>	6 (26%)
<i>Giardia lamblia</i> ( <i>Giardia duodenalis</i> )	7 (25%)
<i>Cryptosporidium</i> spp.	6 (22%)
<i>Shigella</i> /enteroinvasive <i>E. coli</i>	3 (15%)
Shiga toxin-producing <i>E. coli</i>	2 (11%)
<i>E. coli</i> O157	2 (7%)
<i>Vibrio</i> spp.	2 (7%)
<i>Plesiomonas shigelloides</i>	1 (3%)
Sapovirus	1 (3%)
<i>Salmonella</i> spp.	0 (0)
<i>Vibrio cholerae</i>	0 (0)
<i>Yersinia enterocolitica</i>	0 (0)
<i>Cyclospora cayetanensis</i>	0 (0)
<i>Entamoeba histolytica</i>	0 (0)
<i>Clostridium difficile</i> toxin A/B	0 (0)
Adenovirus F 40/41	0 (0)
Astrovirus	0 (0)
Rotavirus	0 (0)

IQR: interquartile range.

which through stool cultures, identified *Salmonella* spp. (mainly *Salmonella enterica*, *Salmonella typhimurium*, and *Salmonella enteritidis*) in first place. However, the isolation of *E. coli*, particularly EAEC, showed it was the pathogen that could contribute to a greater development of diarrheal disease, by reducing local immunity and prolonging disease chronicity<sup>22</sup>. Nevertheless, culture use is still poorly understood, given that it continues to be a test with a questionable yield.

The high level of *E. coli* as a cause of enteric infections in our group of patients may be due to the ingestion of contaminated foods or water<sup>23</sup>, severe immunosuppression, and the practice of unsafe anal sex resulting in fecal-oral exposure. Unsafe anal sex is related to infection with *Campylobacter jejuni*<sup>24</sup>, which was the second most fre-

quent type of bacteria identified by the FilmArray<sup>®</sup> GI Panel.

Said panel detected multiple types of pathogens in one test. In addition to the enteropathogenic bacteria, *E. coli* or *Campylobacter* spp., it identified *Giardia lamblia* as the most frequent parasite, followed by *Cryptosporidium* spp. Our findings differed from those of an Asian study that found *Cryptosporidium* spp. as the main pathogen related to chronic diarrhea in HIV patients, surpassing enteropathogenic bacteria and viruses<sup>25</sup>.

Norovirus was the most frequent viral agent, followed by sapovirus. In a previous Chinese study that utilized the FilmArray<sup>®</sup> GI Panel, norovirus was identified as the main infectious agent in 53% of the study population in patients with HIV and diarrhea, but the etiologic agents were not dif-

**Table 2** Etiologic agent identification results using the FilmArray® GI Panel in patients with HIV infection and chronic diarrhea.

ID	Age	Sex	CD4 (cell/mm <sup>3</sup> )	Viral load (copies/ml)	FilmArray® GI Panel results	Other diagnosis	Treatment regimen/final condition
1	42	M	106	24,500	<i>Campylobacter</i> , <i>Vibrio</i> spp.		Azithromycin, doxycycline/improved
2	39	F	2	36,000	Norovirus		Nitazoxanide/improved
3	35	M	18	197,280	<i>Giardia lamblia</i> , norovirus		Nitazoxanide/improved
4	65	M	12	32,200	None	Intestinal TB through ileocecal biopsy	Isoniazid, rifampicin, ethambutol, pyrazinamide/improved
5	55	M	28	118,000	Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i> Shiga toxin-producing <i>E. coli</i> <i>E. coli</i> O157 <i>Cryptosporidium</i> sp. <i>Giardia lamblia</i> Norovirus		Ciprofloxacin, Nitazoxanide/improved
6	40	M	489	120,000	Enteropathogenic <i>E. coli</i> <i>Giardia lamblia</i>		Ciprofloxacin, Nitazoxanide/improved
7	18	M	138	3,840	<i>Campylobacter</i> spp. Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i>		Ciprofloxacin, azithromycin/improved
8	32	F	204	496,000	Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i> <i>Cryptosporidium</i> sp.		Ciprofloxacin, Nitazoxanide/improved
9	31	M	46	39,400	Enteropathogenic <i>E. coli</i> <i>Giardia lamblia</i>	Intestinal TB through Ziehl-Neelsen stain in stool	(Isoniazid, rifampicin, ethambutol, pyrazinamide), nitazoxanide, ciprofloxacin/improved
10	38	M	2	36,000	Norovirus		Nitazoxanide/improved
11	29	M	44	182,037	Norovirus		Nitazoxanide/improved
12	46	F	189	80,000	None	Histoplasmosis through colonic biopsy	Amphotericin, itraconazole/improved
13	49	M	35	132,000	Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i> Shiga toxin-producing <i>E. coli</i> <i>E. coli</i> O157 <i>Shigella</i> /enteroinvasive <i>E. coli</i>		Ciprofloxacin/improved

Table 2 (Continued)

ID	Age	Sex	CD4 (cell/mm <sup>3</sup> )	Viral load (copies/ml)	FilmArray® GI Panel results	Other diagnosis	Treatment regimen/final condition
14	37	F	131	56,000	Enterotoxigenic <i>E. coli</i> <i>Shigella</i> /enteroinvasive <i>E. coli</i> <i>Cryptosporidium</i> sp. Sapovirus		Ciprofloxacin, Nitazoxanide/improved
15	55	M	28	118,000	<i>Campylobacter</i> spp. Enteroaggregative <i>E. coli</i> Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i> <i>Cryptosporidium</i> sp. <i>Giardia lamblia</i> Norovirus		Azithromycin, Ciprofloxacin, Nitazoxanide/improved
16	19	M	12	25,000	<i>Plesiomonas shigelloides</i> Enteroaggregative <i>E. coli</i> Enteropathogenic <i>E. coli</i> <i>Cryptosporidium</i> sp.		Ciprofloxacin, Nitazoxanide/improved
17	31	M	41	124,700	<i>Campylobacter</i> spp. Enteropathogenic <i>E. coli</i>		Azithromycin, Ciprofloxacin/improved
18	35	F	5	74,000	<i>Campylobacter</i> spp. Enteropathogenic <i>E. coli</i>		Azithromycin, Ciprofloxacin/improved
19	43	F	23	70,800	<i>Vibrio</i> spp. Enteroaggregative <i>E. coli</i> Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i>		Doxycycline, Ciprofloxacin/improved
20	29	M	96	66,300	Enteropathogenic <i>E. coli</i> , <i>Giardia lamblia</i> , Norovirus		Ciprofloxacin, Nitazoxanide/improved
21	23	M	231	20.3	<i>Campylobacter</i> spp. Enteropathogenic <i>E. coli</i> <i>Shigella</i> /enteroinvasive <i>E. coli</i> <i>Cryptosporidium</i> sp. <i>Giardia lamblia</i> Norovirus		Azithromycin, Ciprofloxacin, Nitazoxanide/improved
22	31	M	30	14,400	Enteropathogenic <i>E. coli</i>	Mucormycosis through colonic biopsy	Amphotericin, surgery/died
23	31	M	6	2,510	Enteroaggregative <i>E. coli</i>		Ciprofloxacin/improved
24	72	M	487	20	Enteroaggregative <i>E. coli</i> Enteropathogenic <i>E. coli</i> Enterotoxigenic <i>E. coli</i> Norovirus		Azithromycin, Ciprofloxacin, Nitazoxanide/improved

CD4: CD4 T lymphocyte count; F: female; M: male.

ferentiated according to acute and chronic diarrhea types<sup>26</sup>. Norovirus can produce acute disease, but its chronicity might be due to the GII genotype and coinfection with other viral agents, mainly rotavirus, sapovirus, astrovirus, and adenovirus<sup>27</sup>.

The presence of multiple coexisting pathogens can also create a climate of symbiosis, favoring its permanence, causing effects on the cell transporters that are similar to those caused by the toxins, prolong the destructive and inflammatory effect on the intestinal villi, and produce binding proteins that facilitate adherence of other pathogens in the apical membranes of enterocytes<sup>28,29</sup>.

In our study, two patients had a negative FilmArray® GI Panel result, signifying that it did not detect the entire spectrum of pathogens related to chronic diarrhea in patients with HIV, making it necessary to continue investigating the presence of other pathogens through colonoscopy, mainly in cases of persistent diarrhea.

The limitation of the present study is the fact that the FilmArray® GI Panel does not detect certain opportunistic pathogens in patients with HIV, such as microsporidia, *Cystoisospora belli*, tuberculosis, and histoplasma. Therefore, conventional studies should be performed in parallel with the FilmArray® GI Panel, such as parasite exams with special dyes for detecting tuberculosis and *Cystoisospora belli* (modified Ziehl-Neelsen stain), and patients that continue to present with diarrhea, despite undergoing the treatment regimen, should undergo biopsy of the affected colonic mucosa through colonoscopy. In our study, colonoscopy performed on the patients that did not respond to treatment helped to identify other causes of chronic diarrhea, such as tuberculosis, histoplasma, and mucormycosis.

## Conclusion

Our study results suggest that by utilizing the FilmArray® GI Panel, multiple pathogens associated with chronic diarrhea in patients with HIV can be identified in a single test, facilitating diagnosis and early treatment, and its usefulness in HIV patients justifies its cost. Conventional studies, such as Ziehl-Neelsen staining in stools, should also be maintained. In addition, if chronic diarrhea persists, we recommend studying the patient through colonoscopy for other non-identified agents, such as tuberculosis, histoplasma, and cytomegalovirus.

## Financial disclosure

No specific grants were received from public sector agencies, the business sector, or non-profit organizations in relation to this study.

## Conflict of interest

The authors declare that there is no conflict of interest.

## References

- World Health Organization. Global Health Estimates 2016: deaths by cause, age, sex, by country and by region, 2000–2016. Geneva: World Health Organization; 2018. Available from: [https://www.who.int/healthinfo/global\\_burden\\_disease/estimates/en/index1.html](https://www.who.int/healthinfo/global_burden_disease/estimates/en/index1.html) [Google Scholar].
- Schiller LR, Pardi DS, Sellin JH. Chronic diarrhea: diagnosis and management. *Clin Gastroenterol Hepatol.* 2017;15, <http://dx.doi.org/10.1016/j.cgh.2016.07.028>, 182–193.e3.
- Sánchez TH, Brooks JT, Sullivan PS, et al. Bacterial diarrhea in persons with HIV infection, United States, 1992–2002. *Clin Infect Dis.* 2005;41:1621–7. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/16267735>
- Zingmond DS, Kilbourne AM, Justice AC, et al. Differences in symptom expression in older HIV-positive patients: the veterans aging cohort 3 site study and HIV cost and service utilization study experience. *J Acquir Immune Defic Syndr.* 2003;33 Suppl 2:S84–92. PMID: 12853857.
- Hill A, Balkin A. Risk factors for gastrointestinal adverse events in HIV treated and untreated patients. *AIDS Rev.* 2009;11:30–8. PMID: 19290032.
- MacArthur RD, DuPont HL. Etiology and pharmacologic management of noninfectious diarrhea in HIV-infected individuals in the highly active antiretroviral therapy era. *Clin Infect Dis.* 2012;55:860–7, <http://dx.doi.org/10.1093/cid/cis544>.
- Tian L-G, Wang T-P, Lv S, et al. HIV and intestinal parasite co-infections among a Chinese population: an immunological profile. *Infect Dis Poverty.* 2013;2:18, <http://dx.doi.org/10.1186/2049-9957-2-18>.
- Logan C, Beadsworth MJB, Beeching NJ. HIV and diarrhoea: what is new? *Curr Opin Infect Dis.* 2016;29:486–94, <http://dx.doi.org/10.1097/QCO.0000000000000305>.
- Missaye A, Dagne M, Alemu A, et al. Prevalence of intestinal parasites and associated risk factors among HIV/AIDS patients with pre-ART and on-ART attending dessie hospital ART clinic, Northeast Ethiopia. *AIDS Res Ther.* 2013;10:7, <http://dx.doi.org/10.1186/1742-6405-10-7>.
- Patel TS, Crutchley RD, Tucker AM, et al. Crofelemer for the treatment of chronic diarrhea in patients living with HIV/AIDS. *HIV AIDS (Auckl).* 2013;5:153–62, <http://dx.doi.org/10.2147/HIV.S30948>.
- Montalvo R, Ticona E, Navincopa M, et al. Recurrent diarrhea due to *Cystoisospora belli* in HIV/AIDS patients receiving HAART. *Rev Peru Med Exp Salud Publica.* 2013;30:326–30. PMID: 23949523.
- Machiels JD, Cremers AJH, van Bergen-Verkuyten MCGT, et al. Impact of the BioFire FilmArray gastrointestinal panel on patient care and infection control. *PLoS One.* 2020;15:e0228596, <http://dx.doi.org/10.1371/journal.pone.0228596>, eCollection 2020.
- Hitchcock M, Gómez CA, Banaei N. Low yield of FilmArray GI panel in hospitalized patients with diarrhea: an opportunity for diagnostic stewardship intervention. *J Clin Microbiol.* 2018;56:e01558–1617, <http://dx.doi.org/10.1128/JCM.01558-17>.
- Alejo-Cancho I, Fernández-Avilés F, Capón A, et al. Evaluation of a multiplex panel for the diagnosis of acute infectious diarrhea in immunocompromised hematologic patients. *PLoS One.* 2017;12(November):e0187458, <http://dx.doi.org/10.1371/journal.pone.0187458>.
- Uppal B, Kashyap B, Bhalla P. Enteric pathogens in HIV/AIDS from a tertiary care hospital. *Indian J Community Med.* 2009;34:237–42, <http://dx.doi.org/10.4103/0970-0218.55291>. PMID: 20049303; PMCID: PMC2800905.
- Flórez AC, García DA, Moncada L, et al. Prevalencia de microsporidios y otros parásitos intestinales en pacientes con infección por VIH, Bogotá, 2001. *Biomédica.* 2003;23:274–82, <http://dx.doi.org/10.7705/biomedica.v23i3.1221>.
- Andrade FM, Quiroga A, Builes C, et al. Epidemiología de la infección por el virus de inmunodeficiencia humana en pacientes hospitalizados en una



- institución de alta complejidad y enseñanza universitaria en Medellín, Colombia. *Infectio*. 2016;20:9–16, <http://dx.doi.org/10.1016/j.infect.2015.05.004>.
18. Cello JP, Day LW. Idiopathic AIDS enteropathy and treatment of gastrointestinal opportunistic pathogens. *Gastroenterology*. 2009;136:1952–65, <http://dx.doi.org/10.1053/j.gastro.2008.12.073>.
  19. Buss SN, Leber A, Chapin K, et al. Multicenter Evaluation of the BioFire FilmArray gastrointestinal panel for etiologic diagnosis of infectious gastroenteritis. *J Clin Microbiol*. 2015;53:915–25, <http://dx.doi.org/10.1128/jcm.02674-14>.
  20. Zhan Z, Guo J, Xiao Y, et al. Comparison of BioFire FilmArray gastrointestinal panel versus Luminex xTAG Gastrointestinal Pathogen Panel (xTAG GPP) for diarrheal pathogen detection in China. *Int J Infect Dis*. 2020;99:414–20, <http://dx.doi.org/10.1016/j.ijid.2020.08.020>.
  21. Kreiniz N, Abu-Ahmad A, Golan-Shany O, et al. Shiga-like toxin-producing *Escherichia coli* inducing diarrhea in patients with multiple myeloma diagnosed by BioFire PCR-Film Array. *Acta Haematol*. 2019;142:187–9, <http://dx.doi.org/10.1159/000498906>.
  22. Huang DB, Mohanty A, DuPont HL, et al. A review of an emerging enteric pathogen: enteroaggregative *Escherichia coli*. *J Med Microbiol*. 2006;55:1303–11 <http://www.ncbi.nlm.nih.gov/pubmed/17005776>
  23. Angulo FJ, Swerdlow DL. Bacterial enteric infections in persons infected with human immunodeficiency virus. *Clin Infect Dis*. 1995;21:S84–93 <http://www.ncbi.nlm.nih.gov/pubmed/8547518>
  24. Quinn TC, Goodell SE, Fennell C, et al. Infections with *Campylobacter jejuni* and *Campylobacter*-like organisms in homosexual men. *Ann Intern Med*. 1984;101:187–92 <http://www.ncbi.nlm.nih.gov/pubmed/6547580>
  25. Utami WS, Murhandarwati EH, Artama WT, et al. Cryptosporidium infection increases the risk for chronic diarrhea among people living with HIV in southeast Asia: a systematic review and meta-analysis. *Asia Pac J Public Health*. 2020;32:8–18, <http://dx.doi.org/10.1177/1010539519895422>.
  26. Yang S, Li M, Cheng J, et al. Diagnostic determination of Norovirus infection as one of the major causes of infectious diarrhea in HIV patients using a multiplex polymerase chain reaction assay. *Int J STD AIDS*. 2019;30:550–6, <http://dx.doi.org/10.1177/0956462418824912>.
  27. Lu L, Jia R, Zhong H, et al. Molecular characterization and multiple infections of rotavirus, norovirus, sapovirus, astrovirus and adenovirus in outpatients with sporadic gastroenteritis in Shanghai, China, 2010–2011. *Arch Virol*. 2015;160:1229–38, <http://dx.doi.org/10.1007/s00705-015-2387-1>.
  28. Azer SA, Tuma F, Available from: <https://www.ncbi.nlm.nih.gov/books/NBK544325/>, 2022.
  29. Mathew S, Smatti MK, Al Ansari K, et al. Mixed viral-bacterial infections and their effects on gut microbiota and clinical illnesses in children. *Sci Rep*. 2019;9:865, <http://dx.doi.org/10.1038/s41598-018-37162-w>.